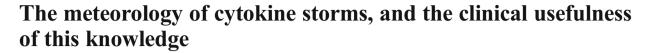
REVIEW



Ian A. Clark¹ · Bryce Vissel^{2,3}

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Abstract The term cytokine storm has become a popular descriptor of the dramatic harmful consequences of the rapid release of polypeptide mediators, or cytokines, that generate inflammatory responses. This occurs throughout the body in both non-infectious and infectious disease states, including the central nervous system. In infectious disease it has become a useful concept through which to appreciate that most infectious disease is not caused directly by a pathogen, but by an overexuberant innate immune response by the host to its presence. It is less widely known that in addition to these roles in disease pathogenesis these same cytokines are also the basis of innate immunity, and in lower concentrations have many essential physiological roles. Here we update this field, including what can be learned through the history of how these interlinking three aspects of biology and disease came to be appreciated. We argue that understanding cytokine storms in their various degrees of acuteness, severity and persistence is essential in order to grasp the pathophysiology of many diseases, and thus the basis of newer therapeutic approaches to treating them. This particularly applies to the neurodegenerative diseases.

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Ian A. Clark ian.clark@anu.edu.au

- ¹ Research School of Biology, Australian National University, Canberra, Australia
- ² School of Life Sciences, Faculty of Science, University of Technology, Sydney, Australia
- ³ Garvan Institute of Medical Research, Sydney, Australia

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Introduction

As previously discussed [1, 2], the term cytokine storm appears to have been first used to describe the chaotic pathophysiological state encountered during an acute graftversus-host disease [3]. The term has caught the popular imagination, with >160,000 hits on a search engine. A flood of harmful polypeptides bursting forth from our cells making us very ill, as if we had severe influenza, is a powerful and accurate image, one clearly worth understanding. Over the intervening 24 years the activity of these cytokines has encompassed a much wider array of biology than dreamed of in 1993, and it is now timely to step back and examine the concept again. In order to provide a wider understanding of the phenomenon of a cytokine storm, we have recounted the history of researchers' awareness of the main mediators whose release initiates the process. Although the popular concept of a cytokine storm is about dramatic illness and disease, these mediators also have essential roles in normal physiology and innate immunity. The principles are the same throughout, but site of production, cytokine concentration and persistence, as well as generation of countering cytokines, determine outcome. The perspectives now possible thus allow us to develop a better understanding of both of these aspects in health and in sickness, as well as focus on the therapeutic possibilities of this knowledge, which is a main theme of this text. We also extend the concept of a cytokine storm into the chronic neurodegenerative states. These diseases remind us that unrelenting moderate rain can destroy, just as surely as a massive storm that causes a flash flood.





Cytokines, the raw material of a cytokine storm

Some 50 years ago the idea of unsuspected secreted polypeptides, mediators akin to hormones but arising throughout the body from cells with other primary functions rather than from specialized secretory organs, and communicating with many other cells, was quite new. The word cytokine became a generic term for these molecules once immunologists realized that classifying them according to their cell of origin, such as lymphocytes releasing lymphokines [4] and monocytes releasing monokines [5] was quite inadequate, particularly since a single mediator could have many cellular origins, as well as pleotropic activity. By 1991, for example, tumor necrosis factor (TNF) had been documented to possess, or at least initiate, 50 biological functions, most demonstrated quite soon after it had been cloned and made available in a recombinant form [6]. TNF [7] is one of the cytokines that kept its original functionbased name. Others include interferons (IFNs) [8] – IFN- γ [9] being particularly relevant – lymphotoxins (LTs) [10] and the transforming growth factors (TGFs) [11]. We should acknowledge here the insight of the authors of a 1979 Letter-to-the-Editor in the Journal of Immunology [12] who proposed the interleukin (IL) form of nomenclature as a way to rescue order out of chaos by suggesting that a number of acronyms describing observed activities could be grouped under a numerical system, beginning with IL-1 and IL-2. This suggestion proved its worth with the increased molecular definition of entities previously known only as activities. Currently IL-1 to IL-37 are recognized, and most, if investigated, are likely to be released in a severe cytokine storm. From what is known of their proinflammatory links to TNF, the cytokines IL-1, IL-12 [13] and IL-17 [14] are particularly likely to contribute to the dramatic outcome of a full-blown storm, but this is the topic of a much wider ranging review. This text is largely restricted to TNF and IL-1, cytokines that conspicuously are released early, and readily induce others. A broad canvas of the array of mediators that can, in excess, form a cytokine storm can be impressive, as can the mathematical models of its dynamics [15], but their known interactions are still in a state of flux. In order to tell the story of the storm rather than the raindrops, we focus here on studies that have contributed to allowing the master initiators, which are released early and set the process going, to be understood, and thus become therapeutic targets.

The biological antiquity of two main players, TNF and IL-1

When a molecule present in mammals proves also to be found in earlier, particularly very early, life forms it is generally taken to denote that it has been conserved in the biological repertoire because it is essential to life. Cytokines can be good examples. For instance, although TNF is large in the mammalian literature it is also found in the marine sponge, *Chondrosia reniformis* [16], and the reef-building corals, *Acropora* spp. [17]. Remarkably, these authors found that coral TNF has a mutual receptor cross-reactivity with human TNF. Similarly, IL-1 is present in the starfish, *Asterias forbesi* [18]. Among insects, cells from at least two genera of moths contain IL-1 and TNF [19], so conceivably all Lepidoptera are similarly endowed. Lower vertebrates such as fish also generate both of these cytokines [20, 21], as well others such as IL-6 [22], IL-8 [23], IL-12 [24] and IL-17 [20].

The discovery of TNF and IL-1 arose from investigating endotoxicity

The earliest published record relevant to this story seems to have been the observation by Maegraith in his 1948 monograph [25] that the functional effects of parenteral bacterial endotoxin, in the form of the typhoid vaccine of the day [26], can be equated with the range of clinical changes observed in falciparum malaria. The clinical non-specificity of malaria, which can mimic rickettsial, bacterial, viral, and noninfectious diseases, carrying with it implications of equal nonspecificity of disease mechanism, has been documented for many decades [27]. Predictably, therefore, whatever mediates the complex illness seen in endotoxicity is acting in these other diseases as well. This conceptual framework has persisted, and continues to be strengthened. The circumstances to which it has now been extended go far beyond its conceptual origins, where gram-negative bacteria, the walls of which contain endotoxin, are present. A crucial next advance was the argument, in 1957, that the pathophysiology of endotoxicity, traumatic shock and hemorrhagic shock are remarkably identical, even to the point of experimentally inducing tolerance to any one of these states protecting against all three [28, 29]. The first arises in bacterial infection, but the other two are decidedly non-infectious.

Although cytokines now loom large in immunology, awareness of them emerged from curiosity about the useful and harmful (endotoxic) effects of bacterial endotoxin. Originally found, as noted above, in cell walls of gramnegative bacteria, endotoxin proved to be a form of lipopolysaccharide (LPS). The term was used interchangeably with LPS for many years, the later taking precedence in recent times. Endotoxin/LPS influences larger life forms in what can seem a bewildering number of ways. Intriguing early phenomena included the in vivo tolerance to its effects observed after a second and subsequent injections [30]. Another major development in 1955 was the capacity of typhoid vaccine, inevitably, we now know, containing LPS, to generate a mediator termed endogenous pyrogen in the serum of rabbits [31]. This was argued to be of leukocyte origin, to be transferable to other rabbits, and not to cross-tolerize with typhoid vaccine [32]. In due course this was one of the paths that led to the recognition of IL-1, as discussed below.

Further complexity arose from the long history of experimental anti-tumor activity [33] of bacterial LPS implying a biological relevance beyond bacterial disease. Others [34] had achieved similar outcomes with an attenuated form of Mycobacterium bovis termed Bacillus Calmette-Guérin (BCG), which greatly sensitizes mice to LPS [35]. Combining these anti-tumor approaches eventually led, in 1975 [7], to the term tumor necrosis factor (TNF), currently with over 135,000 PubMed hits, far more than any other cytokine, entering the scientific lexicon. Until the insights that arose from this work, endotoxin/LPS had been assumed to be directly responsible for its tumorkilling activities. The next year the hypoglycemia induced by LPS [36] proved to arise, as did TNF, through the actions of a macrophage-origin, LPS-induced, polypeptide the authors termed glucocorticoid antagonizing factor (GAF) [37]. As far as its purification was taken, GAF mirrored TNF [38]. These two lines of research pointed the way to appreciating that endotoxin, of itself, is not, as had been thought, directly active against tumors or has the ability to change blood glucose levels, but induces soluble mediators of host origin that have these properties. In other words, mice exposed to parenteral endotoxin generate molecules with anti-tumor and hypoglycemic activity. This opened minds to wondering how many of these mediators existed, and what they did.

Work on the identity of endogenous pyrogen (see [31], above) lay dormant for many years, until 1979, when LPS was appreciated to induce serum amyloid A (SAA) from macrophages indirectly, through the action of another macrophage-origin mediator the authors appropriately termed SAA-inducer [39]. Soon after, this product was found to be identical to both endogenous pyrogen and lymphocyte activating factor (LAF) [40, 41], one of the many names subsumed into the term IL-1 [12]. Hence endogenous pyrogen was evidently identical to IL-1.

What controls TNF increases in innate immunity and disease?

As we have recently reviewed in this context [42], from the late 1980s interest in the evolution of the immune system fostered ideas on how the cytokines that caused innate immunity and disease pathogenesis could arise, and be controlled, in non-infectious as well as infectious disease states. The models that this era generated [43, 44] are still in place. In brief, pathogen-associated molecular patterns (PAMPs) are released by pathogens, and damage-associated molecular patterns (DAMPs) by tissue damaged by trauma, hypoxia, and metals

such as lead, which hypomethylate host DNA. This has the same PAMP activity as do mitochondrial DNA and bacterial DNA, which are innately hypomethylated. Both PAMPs and DAMPs are agonists for non-specific pattern recognition receptors (PPRs) on or inside most cell types, the best described of which are the toll-like receptors, or TLRs [45]. In this way, a disparate collection of signals triggering the same functional outcome fits within a framework that provides these signals with the ability to trigger the release of proinflammatory cytokines. For instance, within this now widespread terminology the LPS discussed in the previous section is one of the PAMPs that are agonists for TLR4 [46]. These cytokines, through the processes of innate immunity, have the capacity to kill the pathogen that provided the PAMP. Also, when in excess, these cytokines initiate pathological processes shared by both infectious and non-infectious diseases.

Some physiological roles of TNF and IL-1 outside the brain

While the principles governing cytokines in innate immunity and disease pathogenesis were being elucidated it was not yet appreciated that TNF and IL-1 are crucial for physiological homeostasis [47]. IL-1 is generally less reported in this context than is TNF, arguably because IL-1-specific reagents are less freely available. Nevertheless, normal hematopoiesis is known to depend on these two cytokines [48, 49], as does normal sleep regulation [50]. Mitochondrial function depends on TNF [51], and it has, as we have reviewed, homeostatic effects on the normal reproduction rate of various progenitor cells, of particular clinical importance those of endothelial cells in severe malaria and sepsis [52]. Clearly, much physiology is controlled by small fluctuations in TNF and IL-1 acting as signaling molecules. The above examples suffice to demonstrate the now established principle that the key cytokines mediating innate immunity and disease pathogenesis are, as next discussed, normally present, and indeed necessarily present, in healthy individuals, fulfilling physiological roles unrelated to inflammation. Thus we advocate not routinely referring to TNF and IL-1 as proinflammatory cytokines, since this terminology often leads this closely linked pair to be regarded simply as biomarkers for the presence of inflammation, a link not made until over 10 years after TNF had been first described [53], and in any event relatively minor within the range of broad biological effects of these cytokines.

Roles of TNF and IL-1 in innate immunity and disease pathogenesis

As has been reviewed [54], in the mid-1970s our laboratory had been seeking a plausible explanation for our observation

that mouse hemoprotozoan parasites die inside circulating erythrocytes during the immune response [55], an unpopular finding in certain quarters, since it was inconsistent with antibody-based dogma on which malaria vaccine research was then based. We were also wondering why a two week prior infection of mice with BCG, administered on the advice of a colleague, Jean-Louis Virelizier, created a milieu inimical to these parasites, causing their death, in the same intraerythrocytic location, some 12 h after their injection into the mice [56]. An additional puzzle was why the illness and pathology caused by these organisms, was, as Maegraith had argued in human malaria decades earlier [25], so similar to endotoxicity. Importantly, we had found we could mimic the mouse disease as well as the death of parasites inside erythrocytes by injecting small amounts of endotoxin/LPS [57]. Remarkably, these changes happened of their own accord in undisturbed self-limiting infections, arguably induced by a stimulus functionally similar to LPS that was provided by the increased parasite load as the infection progressed. Thus we began to search for explanations based on LPS-induced macrophage products that might make sense of these observations. The essential revelation that awakened us to the possibility of TNF being responsible came from reading the then new1975 Sloan Kettering paper [7] on the use of BCG and LPS, reagents we had been using with the same timing, but in our case to understand the relationship between an infectious agent and its host rather than tumor killing. With their collaboration in assaying TNF, we began to develop our then novel view that infectious disease is caused not directly by the invading pathogen, but by what were effectively the side effects of the host's over-exuberant innate immune response to it. We had evidence, albeit now regarded as rudimentary because of the pre-recombinant assays of the day, that TNF and lymphocyte activating factor (LAF), an old name for IL-1, could be involved [58].

This proposal could not, of course, be properly tested until peptides such as cytokines became available in recombinant form, and in sufficient quantity for in vivo use. Since TNF was first recognized for its tumor killing properties, ambitions were high in the late 1980s that rTNF [59] would be therapeutically valuable to cancer patients. In hindsight, we know that severe side-effects [60-62] of the type now attributable to a cytokine storm, were inevitable. To these researchers' minds the side effects of rTNF they produced resembled an influenza-like disease [63], but to us, with our malaria background, they were also profoundly malaria-like. This generally unanticipated outcome of treatment with rTNF was extremely instructive for our theories on the cytokine origins of the illness of malarial disease and bacterial sepsis [58, 64], and we and others had already been injecting rTNF into animal models for this purpose [65, 66]. In the next year, 1988, these principles were incorporated into the original description of sickness behavior [67], which we subsequently described, in its extreme form, as being the basis of the syndrome seen in severe systemic protozoal, bacterial, viral disease, as well as post-trauma [68].

Thus developed the view that infectious disease is caused not directly by the invading pathogen, but by the host's excessive innate immunity to it. The concept eventually spread across the board to include both innate immunity and disease pathogenesis in, for example, infection with *Mycobacterium* spp. [69, 70], *Brucella abortus* [71, 72], *Salmonella* spp. [73, 74], *Listeria monocytogenes* [75, 76], *Leishmania* spp. [77, 78], *Toxoplasma gondii* [79, 80], and influenza [81, 82]. This pattern of cytokines being useful in low concentrations but harmful in high, first reviewed in 1987 in a malaria context [83], is now generally accepted in infectious disease. It is not, however, at all restricted to pathogen-induced conditions, as discussed below.

Many reports link high circulating levels of TNF and IL-1, arguably the original hallmarks of a cytokine storm, with the biochemical details of the pathogenesis of clinical markers of critically ill patients. A random illustration is hypoalbuminemia, an independent marker of poor outcomes in severely ill patients with various diagnoses [84], as recently investigated in critically ill children in intensive care [85]. As reviewed [68], hypoalbuminemia is also a characteristic of malaria, sepsis, acute viral diseases, and severe trauma, all conditions with high TNF. They are, indeed, prototype examples of the effects of a cytokine storm. Reduced albumin is to be expected in all these circumstances, since the liver-specific albumin gene is positively regulated by Dbp [86], one of the circadian genes that TNF and IL-1 suppress [87]. Picomolar concentrations of TNF have been demonstrated to reduce albumin production by human hepatocytes [88]. Whether it does so by inducing IL-1, or independently, or whether they act synergistically, appears, as in too many other circumstances, yet to be determined.

Persistent cytokine storms in the ill brain

Moderate, but persistent, cytokine storms are typical of the chronic neurodegenerative states, including post-stroke, post-traumatic brain injury, and Alzheimer's disease (AD). In the weather analogy, cytokines are what water is to life. Light falls of rain, like low levels of TNF and IL-1, keep physiology ticking over and organisms alive. In moderate amounts rain improves outcome, as does self-limiting innate immunity, but in acute excess, or unrelenting moderate amounts, both rain and cytokines can kill. Both the acute and unrelenting patterns are valid cytokine storms. Acute systemic cytokine excesses, ie those outside the central nervous system (CNS), typically arise from the effects from bacterial or viral PAMPs, and are, if not acutely fatal, generally transient. In contrast, when excess cytokine is generated within the brain in sufficient quantities, as distinct from entering from outside, the usual type of cytokine storm is an unrelenting moderately raised activity that leads to non-resolving inflammation.

This non-resolving pattern of inflammation in the brain is consistent with the consequences of injecting LPS, the prototype TNF inducer, and a TLR4 agonist, in order to generate models of neurodegenerative disease in rodents [89]. Following a single systemic LPS injection, TNF production in mouse brain remains high for at least ten months [90]. In contrast, serum TNF levels peaked at the expected nine hours. Why this difference? These authors argue that an acute systemic injection of LPS activates brain microglia through TNF receptors that initiate sustained activation of brain cytokine synthesis and neuroinflammation. These studies are consistent with earlier work in which the TNF switch-off that occurs systemically after a second LPS injection, which demonstrates the presence of LPS tolerance, proved to be absent in the intracisternal space [91]. Evidently TNF generation is inhibited differently systemically and cerebrally, conceivably mediated either through failure of the anti-inflammatory cytokines IL-4 or IL-10 to increase as they do systemically, and, as noted above, LPS tolerance being weak or absent inside the blood brain barrier (BBB). It is also consistent with the activated state of microglia many years after brain ischemia [92] or brain trauma events [93, 94], as well as with evidence for a positive feedback loop for microglial activation via TNF [95]. Thus the central nervous system is especially vulnerable to cytokine storms that arise when TNF is generated within the brain, from many cell types, particularly microglia and astrocytes but also including neurons [96], and leads to loss of homeostasis in such vulnerable sites as synapses. One predictable consequence is the loss of the subtle homeostasis we depend on for learning, memory, and normal behavior, as seen in chronic neurodegenerative states. As discussed below, severe changes include neuronal death through excitotoxicity.

Since brain function determines subtleties such as personality, behavior, executive function, mood, willpower, learning and memory, we can expect the effects of brain TNF excess to be much more nuanced than the same change in the rest of the body. This is indeed what happens. For example, mice without certain TNF receptors do not become aggressive [97]. The origins of delirium, in which a seriously ill patient shows transient disorientation, confusion and memory loss as part of an exaggerated sickness behavior, remain controversial [98], but it is certainly part of a cytokine storm. It is now considered to be best understood in terms of peripheral TNF being increased sufficiently for enough to cross the BBB for a limited period [99]. Dementia, in contrast, reflects continual TNF production within the brain. Likewise, the coma that is often part of an encephalopathy accompanying sepsis, influenza or malaria can also be rationalized in cytokine terms, with associated coma argued to arise through increased cerebral TNF reducing orexin levels [100]. See reference [101] for a review.

Some key physiological roles of TNF and IL-1 inside the brain

As we have noted [42], physiological roles of TNF and IL-1 inside the brain include their release during physiological neuronal activity and, as has been reviewed [102], playing a crucial role in regulating the strength of normal synaptic transmission. TNF, of itself rather than through the inflammatory cascade it can trigger, is also involved in normal transmission via modulating excitatory neurotransmission [103], trafficking of AMPA receptors [104], homeostatic synaptic scaling [105], long-term potentiation [106], and maintaining normal background levels of neurogenesis [107]. As noted earlier, and of particular relevance in the brain, which requires much oxygen, mitochondrial function depends on TNF [51]. So too does regulation of the neurotransmitter orexin [100], which, as we recently reviewed in a brain disease context [101], controls sleep, motor control, focused mental effort, appetite and water intake. TNF also regulates neuronal type-1 inositol trisphosphate receptors (IP3R), which are central to neuronal Ca⁺⁺ homeostasis, and thus the ionic signaling cascades on which normal function of these cells depends [108]. Likewise, glycine receptors, which are structurally related to γ aminobutyric acid (GABA) receptors and have a similar inhibitory role, are influenced by proinflammatory cytokines [109].

Some key pathophysiological roles of TNF and IL-1 inside the brain

As we have recently reviewed [110], high brain TNF levels increase harmful cerebral glutamate concentrations through enhancing both glutaminase activity and glutamate re-entry proteins, but evidence implies that IL-1 does not [111]. Clearly, these brain functions are susceptible to TNF and/or IL-1 being above their homeostatic range during a cytokine storm, and consequent change can be expected in the subtleties as well as the gross consequences of excitotoxicity in circumstances when the proinflammatory cytokine load is high and prolonged. Neurogenic pain [112] and insulin resistance in AD brains [113] are examples.

Much relevant material on TNF and the brain that space considerations preclude here is contained in our 2010 review [114]. This text developed from our view that the riddle of malarial encephalopathy (cerebral malaria, (CM)) could be understood only by getting engrossed in understanding how TNF influences brain function in other states, such as AD [115]. We felt that in both of these conditions the historical histological hallmarks – sequestered parasitized erythrocytes in CM, and amyloid (A β)plaques in AD – had somehow been allowed, in the absence of other contenders, to evolve into primary mechanisms that researchers were comfortable with, conceivably because they could point them out under a microscope. Increasingly, each hallmark has fallen from the spotlight because of developments in the cytokine storm literature. Importantly, CM has been reported recently to respond dramatically after treating affected mice with the glutamine analog 6-diazo-5-oxo-L-norleucine (DON) [116], lowering the brain glutamate and thus excitotoxicity, that excess brain TNF generates [110].

Likewise, developments in cytokine biology have meant that $A\beta$, for decades almost universally argued to be central to understanding AD, has lost momentum as the key to this disease as well as its proposed importance in post-stroke syndromes [117] and traumatic brain injury [118]. Over two decades ago TNF was reported to alter synaptic transmission in hippocampal slices [119]. Five years later [120] it was shown that this earlier observation explained the ability of $A\beta$, through TNF, to do the same. Other laboratories expanded the roles of TNF in this context [103]. The capacity of $A\beta$ to act as a ligand for CD14 and TLR2 indicates that these findings with A β [120] are consistent with basic immunology, since occupancy of CD14 and TLRs is how the usual bacterial and protozoal-origin inducers of TNF operate [121]. Key support for this concept has been provided by the recent demonstration that the release of pro-inflammatory cytokines from astrocytes is necessary for either A β to be neurotoxic or tau phosphorylation to be initiated [122]. Importantly, a recent large epidemiological study, in which administering regular subcutaneous etanercept, a specific anti-TNF biological agent in common clinical use, over an extended period in treatment of rheumatoid arthritis (RA), was reported to reduce incidence of AD [123] in these patients. This further greatly enhances the likelihood of TNF, and correspondingly decreases the likelihood of $A\beta$, being the key to AD pathogenesis. These and related arguments are discussed in our reviews [42, 52, 110, 124-128].

More recently a technically impressive study [129] has identified changes in the dynamics of re-uptake proteins, and thus of glutamate, in the extracellular microenvironment near A β plaques in the brain of a high A β mouse model of AD. Unfortunately, presenting these data as evidence for a direct functional link between A β and cognitive impairment and neuronal loss through excitotoxicity, as these authors do, ignores the literature on the essential intermediary role of TNF in these observations [120, 130]. The outcome is actually a strong case for AD arising through a chronic cerebral cytokine storm, since TNF, as well as being induced by many TLR agonists, including A β , is well known [110] to influence glutamate dynamics in the way described above [129].

Clinical usefulness of this knowledge

The obvious inference from the literature reviewed here is that patients suffering from cytokine storms should be treated by neutralize the offending cytokines, mainly TNF. It was, however, established quite early in mice [131] and baboons [132], when knowledge of post-TNF pathways was in its infancy, that specific anti-TNF neutralizing agents have to be administered 1-2 h before TNF induction, while animals are still perfectly healthy, in order to save them. An example of a useful application of this principle has been the successful treatment of the life-threatening Jarisch-Herxheimer reaction that can be induced by the PAMPs released from damaged and dead Borrelia duttonii, the cause of African East Coast Relapsing Fever, as a consequence of treating patients with penicillin [133]. Treatment consisted of pre-penicillin exposure to a polyclonal Fab antibody fragments against TNFalpha (anti-TNF-alpha Fab). In contrast, when someone is acutely ill from sepsis the TNF that made them ill has largely come and gone, having set in train many harmful pathways. This is presumably what prevents specific anti-TNF neutralizing agents from being clinically useful in patients who are acutely ill from sepsis [134]. There is also the practical consideration of anti-TNF agents reducing the efficacy of innate immunity in acute infections, albeit first recognized in treatment of RA patients harboring chronic infections such as tuberculosis, in which innate immunity is an important component [135].

Chronic inflammation is quite another story. Some 5 years ago Karl Nathan reviewed the therapeutic challenge represented by resolving compared to non-resolving inflammatory diseases [136]. Specific anti-TNF biologicals have made their mark in a number of these non-resolving inflammatory states, and their influence is set to expand. They are well-established in the treatment of RA, psoriasis and Crohn's disease, three non-resolving conditions affecting different anatomical sites in the periphery. Although for historical reasons these three diseases are the province of three different medical specialties, this has not prevented a common approach to treatment. Moreover, treating the non-resolving inflammatory states that constitute the neurodegenerative diseases by this same approach has, for some time now, awakened great interest in some quarters. Unfortunately, this enthusiasm for treating the brain for excess TNF is, to date, largely restricted to neuroscientists and medical specialists with prior anti-TNF experience in their field [123, 137-139]. It has yet, it seems, to extend to neurologists [128], conceivably in part for commercial-in-confidence reasons [140]. Nevertheless, the value for patients in bridging this knowledge gap has been compelling for some years, particularly with recent awareness that brain TNF levels are a main cause of variation in synaptic activity of glutamate, which, across the neurodegenerative diseases, is manifested when in excess as excitotoxicity

[110]. As discussed [110], this approach gives an important new level of understanding on the use of not only anti-TNF biologicals, but also nilotinab, 6-diazo-5-oxo-norleucine (DON), 3,6 dithio-thalidomides, ceftriaxone, riluzole, and cannabidiol (CBD) in neurodegenerative states. It also explains [141] the capacity of certain stem cells release to improve post-stroke disabilities through creating an antiinflammatory milieu by generating large amounts of fibroblast growth factor rather than by replacing dead cells [142].

Relative importance of TNF and IL-1 as pharmaceutical targets

A useful starting point to discussing this topic is the 1986 report [143] that TNF induces production of IL-1. This is consistent with later studies surrounding the first demonstration of clinical success of infliximab [144], the original specific anti-TNF biological agent, being reported to reduce IL-1 as well as TNF levels [145–147]. It also reduced IL-6 [146, 148], and IL-8 [146]. During these studies this group appears to have been the first to described the concept of a cytokine cascade [146, 148].

Nevertheless, while this literature provides an excellent rationale for the therapeutic use of specific anti-TNF biological agents, there may be an opportunity, particularly regarding IL-1, that is still relatively neglected. There is ample evidence that in certain circumstances IL-1, induced by TNF [143], can itself induce TNF, and the shared and unshared functions of these two cytokines have been discussed at length [149]. These include the membrane-associate IL1 (IL-1 α) [150], as well as circulating IL-1, or IL-1 β [151, 152]. In addition, TNF and IL-1 can synergize [153–155], for example in causing illness and pathology [156, 157]. Thus, although neutralizing excess TNF has become the dominant pharmacological approach in this field, excess IL-1 is receiving, and certainly warrants, further attention as a target molecule.

TNF and IL-1 received roughly equal prominence in the earlier literature, but neutralizing the effects of TNF has largely dominated the therapeutics literature. In part this may be because specific anti-TNF biologicals have been freely available laboratory and clinical tools, since the mid 1980s and mid 1990s respectively. Two specific anti-IL-1ß biologicals, canakinumab [158] and another termed P2D7KK [159], as well as a recombinant form of a naturally occurring antagonist to IL-1 receptor, Anakinra [160, 161], are available. We feel it can fairly be said to date, however, that these agents have proved to be less potent in treating inflammatory disease than are the specific ant-TNF biological agents. In 2013 Dinarello [162] extensively reviewed why this was apparently so. Nevertheless, various applications are still vigorously explored. For example one group has been capitalizing on their finding that IL-1

receptor antagonist (IL-1ra) is an endogenous neuroprotectant that is increased rapidly in a rat model of focal ischemia. This has led them to enhance this antagonist by supplementing its in vivo levels with the recombinant form, Anakinra [163]. They have since reported that, despite a large molecular weight that would preclude its passage through the normal BBB after subcutaneous injection in the rat, Anakinra entered the CSF for a therapeutic window of up to 24 h after the stroke event, and prevented subsequent damage by a reported 33% [164]. This delivery method was also successful after a range of intravenous injection protocols in patients, although clinical outcomes were not measured [165]. An initial controlled trial produced no harm, and was considered too underpowered to achieve positive outcomes [166]. The most likely target for this treatment has been considered to be to increase cerebral blood flow. As noted previously, this group had earlier reported that IL-1ra did not influence glutamate release into synaptosomes [111].

In conclusion, we note that these recent implications of cytokine storms for understanding encephalopathies usefully allow disease pathogenesis to be appreciated as a single entity through bridging the gap between TNF in systemic disease and the brain, as well as encompassing infectious and noninfectious disease on both sides of the blood-brain barrier. This promises new therapeutic perspectives for important cerebral disorders, particularly chronic neurodegenerative states, as exemplified by new data linking cerebral TNF and extracellular glutamate referred to earlier in this text. As we have summarized [110], cerebral TNF and consequentially extracellular cerebral glutamate are both chronically increased in Alzheimer's disease, post-stroke syndromes, traumatic brain injury, and Parkinson's disease, Huntingdon's disease, amylotropic lateral sclerosis, septic encephalopathy, poor post-operative cognition, poor post-irradiation cognition, HIV dementia, cerebral malaria and viral encephalitides. Cerebral palsy also fits this pattern [167, 168]. As noted [110], this relationship between TNF and glutamate is consistent with excitotoxicity and synaptic shutdown being major consequences of chronic cerebral cytokine storms, albeit with different anatomical locations, initiators and kinetics. We suggest that the physiological roles of TNF in the brain are sufficiently diverse to generate, when homeostasis of this cytokine and therefore glutamate are lost, most of the overlapping syndromes encompassed by the above array of disease states. Logical therapeutic approaches include specific anti-TNF or anti-IL-1 agents that enter the CSF, whether through their route of administration [139] or size [110]. With these advances, our understanding of cytokine storms is much closer to coming of age.

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References

- 1. Clark IA (2007) The advent of the cytokine storm. Immunol Cell Biol 85:271–273
- Tisoncik JR, Korth MJ, Simmons CP, Farrar J, Martin TR, Katze MG (2012) Into the eye of the cytokine storm. Microbiol Mol Biol Rev 76(1):16–32. doi:10.1128/mmbr.05015-11
- 3. Ferrara JL (1993) Cytokine dysregulation as a mechanism of graft versus host disease. Curr Opin Immunol 5(5):794–799
- Dumonde DC, Wolstencroft RA, Panayi GS, Matthew M, Morley J, Howson WT (1969) "Lymphokines": non-antibody mediators of cellular immunity generated by lymphocyte activation. Nature 224(5214):38–42
- Kouttab NM, Mehta S, Morgan J, Tannir N, Sahasrabuddhe C, Maizel AL (1984) Lymphokines and monokines as regulators of human lymphoproliferation. Clin Chem 30(9):1539–1545
- Jäättelä M (1991) Biologic activities and mechanisms of action of tumor necrosis factor-a cachectin. Lab Investig 64(6):724–742
- Carswell EA, Old LJ, Kassel RL, Green S, Fiore N, Williamson B (1975) An endotoxin-induced serum factor that causes necrosis of tumors. Proc Natl Acad Sci U S A 72(9):3666–3670
- Isaacs A, Lindenmann J (1957) Virus interference. I. The interferon. Proc R Soc Lond B Biol Sci 147(927):258–267
- Hovanessian AG, La Bonnardiere C, Falcoff E (1980) Action of murine gamma (immune)interferon on beta (fibroblast)-interferon resistant L 1210 and embryonal carcinoma cells. J Interf Res 1(1): 125–135
- Ruddle NH, Waksman BH (1967) Cytotoxic effect of lymphocyte-antigen interaction in delayed hypersensitivity. Science 157(792):1060–1062
- Moses HL, Branum EL, Proper JA, Robinson RA (1981) Transforming growth factor production by chemically transformed cells. Cancer Res 41(7):2842–2848
- 12. Aarden LA (1979) Revised nomenclature for antigen-nonspecific T cell proliferation and helper factors. J Immunol 123:2928–2929
- Gately MK, Desai BB, Wolitzky AG, Quinn PM, Dwyer CM, Podlaski FJ, Familletti PC, Sinigaglia F, Chizonnite R, Gubler U et al (1991) Regulation of human lymphocyte proliferation by a heterodimeric cytokine, IL-12 (cytotoxic lymphocyte maturation factor). J Immunol 147(3):874–882
- Yao Z, Painter SL, Fanslow WC, Ulrich D, Macduff BM, Spriggs MK, Armitage RJ (1995) Human IL-17: a novel cytokine derived from T cells. J Immunol 155(12):5483–5486
- Yiu HH, Graham AL, Stengel RF (2012) Dynamics of a cytokine storm. PLoS One 7(10):e45027. doi:10.1371/journal.pone. 0045027
- Pozzolini M, Scarfi S, Mussino F, Ferrando S, Gallus L, Giovine M (2015) Molecular cloning, characterization, and expression analysis of a prolyl 4-hydroxylase from the marine sponge *Chondrosia reniformis*. Mar Biotechnol (NY) 17(4):393–407. doi:10.1007/s10126-015-9630-3
- Quistad SD, Stotland A, Barott KL, Smurthwaite CA, Hilton BJ, Grasis JA, Wolkowicz R, Rohwer FL (2014) Evolution of TNFinduced apoptosis reveals 550 my of functional conservation. Proc Natl Acad Sci U S A 111(26):9567–9572. doi:10.1073/pnas. 1405912111

- Beck G, Habicht GS (1986) Isolation and characterization of a primitive interleukin-1-like protein from an invertebrate, *Asterias forbesi*. Proc Natl Acad Sci U S A 83(19):7429–7433
- Wittwer D, Franchini A, Ottaviani E, Wiesner A (1999) Presence of IL-1- and TNF-like molecules in galleria mellonella (Lepidoptera) haemocytes and in an insect cell line Fromestigmene acraea (Lepidoptera). Cytokine 11(9):637–642. doi:10.1006/cyto.1998.0481
- Gunimaladevi I, Savan R, Sakai M (2006) Identification, cloning and characterization of interleukin-17 and its family from zebrafish. Fish Shellfish Immunol 21(4):393–403. doi:10.1016/j. fsi.2006.01.004
- Wang T, Bird S, Koussounadis A, Holland JW, Carrington A, Zou J, Secombes CJ (2009) Identification of a novel IL-1 cytokine family member in teleost fish. J Immunol 183(2):962–974. doi: 10.4049/jimmunol.0802953
- Wang T, Secombes CJ (2009) Identification and expression analysis of two fish-specific IL-6 cytokine family members, the ciliary neurotrophic factor (CNTF)-like and M17 genes, in rainbow trout *Oncorhynchus mykiss*. Mol Immunol 46(11–12):2290–2298. doi: 10.1016/j.molimm.2009.04.003
- Wang E, Wang J, Long B, Wang K, He Y, Yang Q, Chen D, Geng Y, Huang X, Ouyang P, Lai W (2016) Molecular cloning, expression and the adjuvant effects of interleukin-8 of channel catfish (*Ictalurus punctatus*) against streptococcus iniae. Sci Rep 6: 29310. doi:10.1038/srep29310
- Wang T, Husain M, Hong S, Holland JW (2014) Differential expression, modulation and bioactivity of distinct fish IL-12 isoforms: implication towards the evolution of Th1-like immune responses. Eur J Immunol 44(5):1541–1551. doi:10.1002/eji. 201344273
- 25. Maegraith B (1948) Pathological processes in malaria and Blackwater fever. In. Blackwell, Oxford, pp 367–369
- Altschule MD, Freedberg AS, McManus MJ (1945) Circulation and respiration during an episode of chill and fever in man. J Clin Invest 24(6):878–889
- Hyman AS (1945) Clinical masquerades of malaria. Observations in South Pacific combat areas. US Naval Medical Bulletin 45: 287–308
- Zweifach BW, Benacerraf B, Thomas L (1957) The relationship between the vascular manifestations of shock produced by endotoxin, trauma, and hemorrhage. II. The possible role of the reticulo-endothelial system in resistance to each type of shock. J Exp Med 106:403–414
- Zweifach BW, Thomas L (1957) The relationship between the vascular manifestations of shock produced by endotoxin, trauma, and hemorrhage. I. Certain similarities between the reactions in normal and endotoxin-tolerant rats. J Exp Med 106(3):385–401
- Biozzi G, Benacerraf B, Halpern BN (1955) The effect of Salm. Typhi and its endotoxin on the phagocytic activity of the reticuloendothelial system in mice. Br J Exp Pathol 36(3):226–235
- Atkins E, Wood WB Jr (1955) Studies on the pathogenesis of fever. I. The presence of transferable pyrogen in the blood stream following the injection of typhoid vaccine. J Exp Med 101(5): 519–528
- Atkins E, Wood WB Jr (1955) Studies on the pathogenesis of fever. II. Identification of an endogenous pyrogen in the blood stream following the injection of typhoid vaccine. J Exp Med 102(5):499–516
- Prigal SJ (1961) Development in mice of prolonged non-specific resistance to sarcoma implant and staphylococcus infection following repository injection of lipopolysaccharide. Nature 191: 1111–1112
- Old LJ, Clarke DA, Benacerraf B (1959) Effect of bacillus Calmette Guérin infection on transplanted tumours in the mouse. Nature 184(4682):291–292

- 35. Suter E (1964) Hyperreactivity to endotoxin after infection with BCG. J Immunol 92:49–54
- Shands JW Jr, Miller V, Martin H (1969) The hypoglycemic activity of endotoxin. I. Occurrence in animals hyperreactive to endotoxin. Proc Soc Exp Biol Med 130(2):413–417
- Moore RN, Goodrum KJ, Berry LJ (1976) Mediation of an endotoxic effect by macrophages. J Reticuloendothel Soc 19(3):187– 197
- Hill MR, Mccallum RE (1992) Identification of tumor necrosis factor as a transcriptional regulator of the phosphoenolpyruvate carboxykinase gene following endotoxin treatment of mice. Infect Immun 60(10):4040–4050
- Sipe JD, Vogel SN, Ryan JL, McAdam KP, Rosenstreich DL (1979) Detection of a mediator derived from endotoxinstimulated macrophages that induces the acute phase serum amyloid a response in mice. J Exp Med 150(3):597–606
- 40. Sztein MB, Vogel SN, Sipe JD, Murphy PA, Mizel SB, Oppenheim JJ, Rosenstreich DL (1981) The role of macrophages in the acute-phase response: SAA inducer is closely related to lymphocyte activating factor and endogenous pyrogen. Cell Immunol 63(1):164–176
- 41. McAdam KP, Li J, Knowles J, Foss NT, Dinarello CA, Rosenwasser LJ, Selinger MJ, Kaplan MM, Goodman R, Herbert PN, Bausserman LL, Nadler LM (1982) The biology of SAA: identification of the inducer, in vitro synthesis, and heterogeneity demonstrated with monoclonal antibodies. Ann N Y Acad Sci 389:126–136
- 42. Clark IA, Vissel B (2015) Amyloid beta: one of three dangerassociated molecules that are secondary inducers of the proinflammatory cytokines that mediate Alzheimer's disease. Br J Pharmacol 172(15):3714–3727. doi:10.1111/bph.13181
- Janeway CA Jr (1989) Pillars article: approaching the asymptote? Evolution and revolution in immunology. Cold Spring Harb Symp Quant Biol 54(9):1–13
- Matzinger P (2002) The danger model: a renewed sense of self. Science 296(5566):301–305
- 45. Poltorak A, Smirnova I, He X, Liu MY, Van Huffel C, McNally O, Birdwell D, Alejos E, Silva M, Du X, Thompson P, Chan EK, Ledesma J, Roe B, Clifton S, Vogel SN, Beutler B (1998) Genetic and physical mapping of the Lps locus: identification of the toll-4 receptor as a candidate gene in the critical region. Blood Cells Mol Dis 24(3):340–355
- 46. Hoshino K, Takeuchi O, Kawai T, Sanjo H, Ogawa T, Takeda Y, Takeda K, Akira S (1999) Cutting edge: toll-like receptor 4 (TLR4)-deficient mice are hyporesponsive to lipopolysaccharide: evidence for TLR4 as the Lps gene product. J Immunol 162(7): 3749–3752
- Tovey MG (1988) The expression of cytokines in the organs of normal individuals: role in homeostasis. A review. J Biol Regul Homeost Agents 2(2):87–92
- Clark IA, Chaudhri G (1988) Tumour necrosis factor may contribute to the anaemia of malaria by causing dyserythropoiesis and erythrophagocytosis. Br J Haematol 70(1):99–103
- Bagby GC Jr (1989) Interleukin-1 and hematopoiesis. Blood Rev 3(3):152–161
- Jewett KA, Krueger JM (2012) Humoral sleep regulation; interleukin-1 and tumor necrosis factor. Vitam Horm 89:241– 257. doi:10.1016/b978-0-12-394623-2.00013-5
- 51. Sanchez-Alcazar JA, Schneider E, Martinez MA, Carmona P, Hernandez-Munoz I, Siles E, De la Torre P, Ruiz-Cabello J, Garcia I, Solis-Herruzo JA (2000) Tumor necrosis factor-alpha increases the steady-state reduction of cytochrome b of the mitochondrial respiratory chain in metabolically inhibited L929 cells. J Biol Chem 275(18):13353–13361
- Clark I, Atwood C, Bowen R, Paz-Filho G, Vissel B (2012) Tumor necrosis factor-induced cerebral insulin resistance in Alzheimer's

disease links numerous treatment rationales. Pharmacol Rev 64(4):1004–1026. doi:10.1124/pr.112.005850

- Oppenheim JJ, Lew W, Akahoshi T, Matsushima K, Neta R (1988) Aspects of cytokine induced modulation of immunity and inflammation with emphasis on interleukin 1. Arzneimittel -Forschung/Drug Research 38(3A):461–465
- Clark IA (2007) How TNF was recognized to be a key mechanism of disease. Cytokine Growth FR 18:335–343
- Clark IA, Richmond JE, Wills EJ, Allison AC (1977) Intraerythrocytic death of the parasite in mice recovering from infection with *Babesia microti*. Parasitology 75(2):189–196
- Clark IA, Wills EJ, Richmond JE, Allison AC (1977) Suppression of babesiosis in BCG-infected mice and its correlation with tumor inhibition. Infect Immun 17(2):430–438
- Clark IA (1978) Does endotoxin cause both the disease and parasite death in acute malaria and babesiosis? Lancet ii 8080:75–77
- Clark IA, Virelizier J-L, Carswell EA, Wood PR (1981) Possible importance of macrophage-derived mediators in acute malaria. Infect Immun 32(3):1058–1066
- Aggarwal BB, Kohr WJ, Hass PE, Moffat B, Spencer SA, Henzel WJ, Bringman TS, Nedwin GE, Goeddel DV, Harkins RN (1985) Human tumor necrosis factor: production, purification, and characterization. J Biol Chem 260(4):2345–2354
- Spriggs DR, Sherman ML, Kufe DW, Frei E (1987) Tumour necrosis factor: clinical trials and future directions. In: Tumour Necrosis Factor and Related Cytokines, Ciba Foundation Symposium, John Wiley and Sons, Chichester, pp 206–227
- Spriggs DR, Sherman ML, Michie H, Arthur KA, Imamura K, Wilmore D, Frei E, Kufe DW (1988) Recombinant human tumor necrosis factor administered as a 24-hour intravenous infusion. A phase 1 and pharmacologic study. J Natl Cancer Inst 80:1039– 1044
- 62. Sherman ML, Spriggs DR, Arthur KA, Imamura K, Frei EF, Kufe DW (1988) Recombinant human tumor necrosis factor administered as a five-day continuous infusion in cancer patients: phase 1 toxicity and effects on lipid metabolism. J Clin Oncol 6:344–350
- Demetri GD, Spriggs DR, Sherman ML, Arthur KA, Imamura K, Kufe DW (1989) A phase I trial of recombinant human tumor necrosis factor and interferon-gamma: effects of combination cytokine administration in vivo. J Clin Oncol 7(10):1545–1553
- Clark IA (1982) Suggested importance of monokines in pathophysiology of endotoxin shock and malaria. Klin Wochenschr 60(14):756–758
- Tracey KJ, Beutler B, Lowry SF, Merryweather J, Wolpe S, Milsark IW, Hariri RJ, Fahey TJ, Zentella A, Albert JD, Shires GT, Cerami A (1986) Shock and tissue injury induced by recombinant human cachectin. Science 234(4775):470–474
- Clark IA, Cowden WB, Butcher GA, Hunt NH (1987) Possible roles of tumor necrosis factor in the pathology of malaria. Am J Pathol 129(1):192–199
- 67. Hart BL (1988) Biological basis of the behavior of sick animals. Neurosci Biobehav Rev 12(2):123–137
- Clark IA, Budd AC, Alleva LM (2008) Sickness behaviour pushed too far–the basis of the syndrome seen in severe protozoal, bacterial and viral diseases and post-trauma. Malar J 7:208
- Bermudez LE, Young LS (1988) TNF, alone or in combination with IL-2, but not IFN-gamma, is associated with macrophage killing of *Mycobacterium avium* Complex. J Immunol 140(9): 3006–3013
- Rook GAW (1987) Progress in the immunology of the mycobacterioses. Clin Exp Immunol 69:1–9
- Zhan YF, Liu ZQ, Cheers C (1996) Tumor necrosis factor alpha and interleukin-12 contribute to resistance to the intracellular bacterium *Brucella abortus* by different mechanisms. Infect Immun 64(7):2782–2786

- Ahmed K, Al Matrouk KA, Martinez G, Oishi K, Rotimi VO, Nagatake T (1999) Increased serum levels of interferon-gamma and interleukin-12 during human brucellosis. AmJTrop Med Hyg 61(3):425–427
- Degre M, Bukholm G (1990) Effect of tumor necrosis factor-alpha on infection with *Salmonella typhimurium* in a mouse model. J Biol Regul Homeost Agents 4(4):157–161
- Bhutta ZA, Mansoorali N, Hussain R (1997) Plasma cytokines in paediatric typhoidal salmonellosis: correlation with clinical course and outcome. J Inf Secur 35(3):253–256
- 75. Rothe J, Lesslauer W, Lotscher H, Lang Y, Koebel P, Kontgen F, Althage A, Zinkernagel R, Steinmetz M, Bluethmann H (1993) Mice lacking the tumour necrosis factor receptor-1 are resistant to TNF-mediated toxicity but highly susceptible to infection by *Listeria monocytogenes*. Nature 364(6440):798–802
- Nakane M, Klinghofer V, Kuk JE, Donnelly JL, Budzik GP, Pollock JS, Basha F, Carter GW (1995) Novel potent and selective inhibitors of inducible nitric oxide synthase. Molec Pharmacol 47(4):831–834
- Titus RG, Sherry B, Cerami A (1989) Tumor necrosis factor plays a protective role in experimental murine cutaneous leishmaniasis. J Exp Med 170:2097–2104
- Raziuddin S, Abdalla RE, el Awad EH, al Janadi M (1994) Immunoregulatory and proinflammatory cytokine production in visceral and cutaneous leishmaniasis. J Infect Dis 170(4):1037– 1040
- Chang HR, Grau GE, Perchere JC (1990) Role of TNF and IL-1 in infections with *Toxoplasma gondii*. Immunol 69:33–37
- Arsenijevic D, Girardier L, Seydoux J, Chang HR, Dulloo AG (1997) Altered energy balance and cytokine gene expression in a murine model of chronic infection with *Toxoplasma gondii*. Am J Phys 272(5):E908–E917
- Sladkova T, Kostolansky F (2006) The role of cytokines in the immune response to influenza a virus infection. Acta Virol 50(3): 151–162
- Hussell T, Pennycook A, Openshaw PJ (2001) Inhibition of tumor necrosis factor reduces the severity of virus-specific lung immunopathology. Eur J Immunol 31(9):2566–2573. doi:10.1002/ 1521-4141(200109)31:9<2566::aid-immu2566>3.0.co; 2-1
- Clark IA (1987) Cell-mediated immunity in protection and pathology of malaria. Parasitol Today 3(10):300–305
- Goldwasser P, Feldman J (1997) Association of serum albumin and mortality risk. J Clin Epidemiol 50(6):693–703
- Kittisakmontri K, Reungrongrat S, Lao-Araya M (2016) Hypoalbuminaemia at admission predicts the poor outcomes in critically ill children. Anaesthesiol Intensive Ther 48(3):158–161. doi:10.5603/AIT.a2016.0028
- Cereghini S (1996) Liver-enriched transcription factors and hepatocyte differentiation. FASEB J 10(2):267–282
- Cavadini G, Petrzilka S, Kohler P, Jud C, Tobler I, Birchler T, Fontana A (2007) TNF alpha suppresses the expression of clock genes by interfering with E-box-mediated transcription. Proc Natl Acad Sci U S A 104(31):12843–12848
- Perlmutter DH, Dinarello CA, Punsal PI, Colten HR (1986) Cachectin/tumor necrosis factor regulates hepatic acute-phase gene expression. J Clin Invest 78(5):1349–1354
- Hauss-Wegrzyniak B, Dobrzanski P, Stoehr JD, Wenk GL (1998) Chronic neuroinflammation in rats reproduces components of the neurobiology of Alzheimer's disease. Brain Res 780(2):294–303
- Qin B, Qiu W, Avramoglu RK, Adeli K (2007) Tumor necrosis factor-alpha induces intestinal insulin resistance and stimulates the overproduction of intestinal apolipoprotein B48-containing lipoproteins. Diabetes 56(2):450–461

- Steinshamn S, Waage A (2000) Lack of endotoxin tolerance with respect to TNF alpha production in the subarachnoid space. APMIS 108(2):107–112
- 92. Pappata S, Levasseur M, Gunn RN, Myers R, Crouzel C, Syrota A, Jones T, Kreutzberg GW, Banati RB (2000) Thalamic microglial activation in ischemic stroke detected in vivo by PET and [11C]PK1195. Neurology 55(7):1052–1054
- 93. Ramlackhansingh AF, Brooks DJ, Greenwood RJ, Bose SK, Turkheimer FE, Kinnunen KM, Gentleman S, Heckemann RA, Gunanayagam K, Gelosa G, Sharp DJ (2011) Inflammation after trauma: microglial activation and traumatic brain injury. Ann Neurol 70(3):374–383. doi:10.1002/ana.22455
- Johnson VE, Stewart JE, Begbie FD, Trojanowski JQ, Smith DH, Stewart W (2013) Inflammation and white matter degeneration persist for years after a single traumatic brain injury. Brain 136(Pt 1):28–42. doi:10.1093/brain/aws322
- Kuno R, Wang J, Kawanokuchi J, Takeuchi H, Mizuno T, Suzumura A (2005) Autocrine activation of microglia by tumor necrosis factor-alpha. J Neuroimmunol 162(1–2):89–96. doi:10. 1016/j.jneuroim.2005.01.015
- Liu T, Clark RK, Mcdonnell PC, Young PR, White RF, Barone FC, Feuerstein GZ (1994) Tumor necrosis factor-alpha expression in ischemic neurons. Stroke 25(7):1481–1488
- Patel A, Siegel A, Zalcman SS (2010) Lack of aggression and anxiolytic-like behavior in TNF receptor (TNF-R1 and TNF-R2) deficient mice. Brain Behav Immun 24(8):1276–1280
- Cunningham C, Maclullich AM (2013) At the extreme end of the psychoneuroimmunological spectrum: delirium as a maladaptive sickness behaviour response. Brain Behav Immun 28:1–13. doi: 10.1016/j.bbi.2012.07.012
- Fong TG, Davis D, Growdon ME, Albuquerque A, Inouye SK (2015) The interface between delirium and dementia in elderly adults. Lancet Neurol 14(8):823–832. doi:10.1016/s1474-4422(15)00101-5
- 100. Zhan S, Cai GQ, Zheng A, Wang Y, Jia J, Fang H, Yang Y, Hu M, Ding Q (2011) Tumor necrosis factor-alpha regulates the hypocretin system via mRNA degradation and ubiquitination. Biochim Biophys Acta 1812(4):565–571. doi:10.1016/j.bbadis. 2010.11.003
- Clark IA, Vissel B (2014) Inflammation-sleep interface in brain disease: TNF, insulin, orexin. J Neuroinflammation 11:51. doi:10. 1186/1742-2094-11-51
- Marin I, Kipnis J (2013) Learning and memory and the immune system. Learn Mem 20(10):601–606. doi:10.1101/lm.028357.112
- Pickering M, Cumiskey D, O'Connor JJ (2005) Actions of TNFalpha on glutamatergic synaptic transmission in the central nervous system. Exp Physiol 90(5):663–670
- 104. Ferguson AR, Christensen RN, Gensel JC, Miller BA, Sun F, Beattie EC, Bresnahan JC, Beattie MS (2008) Cell death after spinal cord injury is exacerbated by rapid TNFalpha-induced trafficking of GluR2-lacking AMPARs to the plasma membrane. J Neurosci 28(44):11391–11400
- Stellwagen D, Malenka RC (2006) Synaptic scaling mediated by glial TNF-alpha. Nature 440(7087):1054–1059
- 106. Cumiskey D, Butler MP, Moynagh PN, O'Connor JJ (2007) Evidence for a role for the group I metabotropic glutamate receptor in the inhibitory effect of tumor necrosis factor-alpha on longterm potentiation. Brain Res 1136(1):13–19
- Bernardino L, Agasse F, Silva B, Ferreira R, Grade S, Malva JO (2008) Tumor necrosis factor-alpha modulates survival, proliferation, and neuronal differentiation in neonatal subventricular zone cell cultures. Stem Cells 26(9):2361–2371
- Park KM, Yule DI, Bowers WJ (2008) Tumor necrosis factoralpha potentiates intraneuronal CA2+ signaling via regulation of the inositol 1,4,5-trisphosphate receptor. J Biol Chem 283:33069– 33079

- Chirila AM, Brown TE, Bishop RA, Bellono NW, Pucci FG, Kauer JA (2014) Long-term potentiation of glycinergic synapses triggered by interleukin 1beta. Proc Natl Acad Sci U S A 111(22): 8263–8268. doi:10.1073/pnas.1401013111
- 110. Clark IA, Vissel B (2016) Excess cerebral TNF causing glutamate excitotoxicity rationalizes treatment of neurodegenerative diseases and neurogenic pain by anti-TNF agents. J Neuroinflammation 13(1):236. doi:10.1186/s12974-016-0708-2
- 111. Allan SM, Lawrence CB, Rothwell NJ (1998) Interleukin-1 beta and interleukin-1 receptor antagonist do not affect glutamate release or calcium entry in rat striatal synaptosomes. Mol Psychiatry 3(2):178–182
- 112. Sommer C, Lindenlaub T, Teuteberg P, Schafers M, Hartung T, Toyka KV (2001) Anti-TNF-neutralizing antibodies reduce painrelated behavior in two different mouse models of painful mononeuropathy. Brain Res 913(1):86–89
- Liu Y, Liu F, Grundke Iqbal I, Iqbal K, Gong CX (2011) Deficient brain insulin signalling pathway in Alzheimer's disease and diabetes. J Pathol 225(1):54–62
- 114. Clark IA, Alleva LM, Vissel B (2010) The roles of TNF in brain dysfunction and disease. Pharmacol Ther 128:519–548
- Clark IA, Alleva LM (2009) Is human malarial coma caused, or merely deepened, by sequestration? Trends Parasitol 25(7):314– 318
- 116. Gordon GR, Baimoukhametova DV, Hewitt SA, Rajapaksha WR, Fisher TE, Bains JS (2005) Norepinephrine triggers release of glial ATP to increase postsynaptic efficacy. Nat Neurosci 8(8):1078– 1086
- 117. Thiel A, Cechetto DF, Heiss WD, Hachinski V, Whitehead SN (2014) Amyloid burden, neuroinflammation, and links to cognitive decline after ischemic stroke. Stroke 45(9):2825–2829. doi: 10.1161/strokeaha.114.004285
- Acosta SA, Tajiri N, Sanberg PR, Kaneko Y, Borlongan CV (2016) Increased amyloid precursor protein and tau expression manifests as key secondary cell death in chronic traumatic brain injury. J Cell Physiol. doi:10.1002/jcp.25629
- 119. Tancredi V, D'Arcangelo G, Grassi F, Tarroni P, Palmieri G, Santoni A, Eusebi F (1992) Tumor necrosis factor alters synaptic transmission in rat hippocampal slices. Neurosci Lett 146(2):176– 178
- Rowan MJ, Klyubin I, Wang Q, Hu NW, Anwyl R (2007) Synaptic memory mechanisms: Alzheimer's disease amyloid beta-peptide-induced dysfunction. Biochem Soc Trans 35(Pt 5): 1219–1223
- Beutler B, Poltorak A (2001) Sepsis and evolution of the innate immune response. Crit Care Med 29(7 Suppl S):S2–S6
- 122. Garwood CJ, Pooler AM, Atherton J, Hanger DP, Noble W (2011) Astrocytes are important mediators of Abeta-induced neurotoxicity and tau phosphorylation in primary culture. Cell Death Dis 2011(2):e167
- 123. Chou RC, Kane M, Ghimire S, Gautam S, Gui J (2016) Treatment for rheumatoid arthritis and risk of Alzheimer's disease: a nested case-control analysis. CNS Drugs. doi:10.1007/s40263-016-0374-z
- Clark IA, Alleva LM, Vissel B (2011) TNF and leptin tell essentially the same story in Alzheimer's disease. J Alzheimers Dis 26(2):201–205
- 125. Clark IA, Vissel B (2013) Treatment implications of the altered cytokine-insulin axis in neurodegenerative disease. Biochem Pharmacol 86:862–871. doi:10.1016/j.bcp.2013.07.030
- 126. Wright AL, Zinn R, Hohensinn B, Konen LM, Beynon SB, Tan RP, Clark IA, Abdipranoto A, Vissel B (2013) Neuroinflammation and neuronal loss precede Abeta plaque deposition in the hAPP-J20 mouse model of Alzheimer's disease. PLoS One 8(4):e59586. doi:10.1371/journal.pone.0059586

- 127. Morris GP, Clark IA, Vissel B (2014) Inconsistencies and controversies surrounding the amyloid hypothesis of Alzheimer's disease. Acta Neuropathol Commun 2:135. doi:10.1186/s40478-014-0135-5
- Clark IA, Vissel B (2015) A neurologist's guide to TNF biology and to the principles behind the therapeutic removal of excess TNF in disease. Neural Plast 2015:358263. doi:10.1155/2015/ 358263
- 129. Hefendehl JK, LeDue J, Ko RW, Mahler J, Murphy TH, MacVicar BA (2016) Mapping synaptic glutamate transporter dysfunction in vivo to regions surrounding Abeta plaques by iGluSnFR twophoton imaging. Nat Commun 7:13441. doi:10.1038/ ncomms13441
- Wang QW, Wu JQ, Rowan MJ, Anwyl R (2005) Beta-amyloid inhibition of long-term potentiation is mediated via tumor necrosis factor. Eur J Neurosci 22(11):2827–2832
- Beutler B, Milsark IW, Cerami AC (1985) Passive immunization against cachectin/tumor necrosis factor protects mice from lethal effects of endotoxin. Science 229(4716):869–871
- 132. Tracey KJ, Fong Y, Hesse DG, Manogue KR, Lee AT, Kuo GC, Lowry SF, Cerami A (1987) Anti-cachectin/TNF monoclonal antibodies prevent septic shock during lethal shock bacteraemia. Nature 330(6149):662–664
- 133. Fekade D, Knox K, Hussein K, Melka A, Lalloo DG, Coxon RE, Warrell DA (1996) Prevention of Jarisch-Herxheimer reactions by treatment with antibodies against tumor necrosis factor alpha. N Engl J Med 335(5):311–315
- 134. Abraham E, Anzueto A, Gutierrez G, Tessler S, Sanpedro G, Wunderink R, Dalnogare A, Nasraway S, Berman S, Cooney R, Levy H, Baughman R, Rumbak M, Light RB, Poole L, Allred R, Constant J, Pennington J, Porter S (1998) Double-blind randomised controlled trial of monoclonal antibody to human tumour necrosis factor in treatment of septic shock. Lancet 351(9107):929–933
- 135. Keane J, Gershon S, Wise RP, Mirabile-Levens E, Kasznica J, Schwieterman WD, Siegel JN, Braun MM (2001) Tuberculosis associated with infliximab, a tumor necrosis factor alphaneutralizing agent. N Engl J Med 345(15):1098–1104. doi:10. 1056/NEJMoa011110
- Nathan C, Ding A (2010) Nonresolving inflammation. Cell 140(6):871–882. doi:10.1016/j.cell.2010.02.029
- 137. Tobinick EL, Gross H, Weinberger A, Cohen H (2006) TNF-alpha modulation for treatment of Alzheimer's disease: a 6- month pilot study. MedGenMed Neurol Neurosurg 8(2):25
- 138. Tobinick E, Kim NM, Reyzin G, Rodriguez-Romanacce H, DePuy V (2012) Selective TNF inhibition for chronic stroke and traumatic brain injury: an observational study involving 629 consecutive patients treated with perispinal etanercept. CNS Drugs 26(12):1051–1070. doi:10.1007/s40263-012-0013-2
- Ignatowski TA, Spengler RN, Dhandapani KM, Folkersma H, Butterworth RF, Tobinick E (2014) Perispinal etanercept for post-stroke neurological and cognitive dysfunction: scientific rationale and current evidence. CNS Drugs 28:679–697. doi:10. 1007/s40263-014-0174-2
- Sumbria RK, Boado RJ, Pardridge WM (2012) Brain protection from stroke with intravenous TNFalpha decoy receptor-Trojan horse fusion protein. J Cereb Blood Flow Metab 32(10):1933– 1938. doi:10.1038/jcbfm.2012.97
- 141. Clark IA (2016) Letter by Clark regarding article, "clinical outcomes of transplanted modified bone marrow-derived mesenchymal stem cells in stroke: a phase 1/2a study". Stroke 47(12):e268. doi:10.1161/strokeaha.116.014920
- 142. Steinberg GK, Kondziolka D, Wechsler LR, Lunsford LD, Coburn ML, Billigen JB, Kim AS, Johnson JN, Bates D, King B, Case C, McGrogan M, Yankee EW, Schwartz NE (2016) Clinical outcomes of transplanted modified bone marrow-derived

mesenchymal stem cells in stroke: a phase 1/2a study. Stroke 47(7):1817–1824. doi:10.1161/strokeaha.116.012995

- 143. Dinarello CA, Cannon JG, Wolff SM, Bernheim HA, Beutler B, Cerami A, Figari IS, Palladino MA, O'Connor JV (1986) Tumor necrosis factor (cachectin) is an endogenous pyrogen and induces production of interleukin-I. J Exp Med 163:1433–1450
- 144. Elliott MJ, Maini RN, Feldmann M, Kalden JR, Antoni C, Smolen JS, Leeb B, Breedveld FC, Macfarlane JD, Bijl H et al (1994) Randomised double-blind comparison of chimeric monoclonal antibody to tumour necrosis factor alpha (cA2) versus placebo in rheumatoid arthritis. Lancet 344(8930):1105–1110
- 145. Brennan FM, Chantry D, Jackson A, Maini R, Feldmann M (1989) Inhibitory effect of TNF alpha antibodies on synovial cell interleukin-1 production in rheumatoid arthritis. Lancet 2(8657): 244–247
- 146. Butler DM, Maini RN, Feldmann M, Brennan FM (1995) Modulation of proinflammatory cytokine release in rheumatoid synovial membrane cell cultures. Comparison of monoclonal anti TNF-alpha antibody with the interleukin-1 receptor antagonist. Eur Cytokine Netw 6(4):225–230
- 147. Lorenz HM, Antoni C, Valerius T, Repp R, Grunke M, Schwerdtner N, Nusslein H, Woody J, Kalden JR, Manger B (1996) In vivo blockade of TNF-alpha by intravenous infusion of a chimeric monoclonal TNF-alpha antibody in patients with rheumatoid arthritis. Short term cellular and molecular effects. J Immunol 156(4):1646–1653
- 148. Charles P, Elliott MJ, Davis D, Potter A, Kalden JR, Antoni C, Breedveld FC, Smolen JS, Eberl G, deWoody K, Feldmann M, Maini RN (1999) Regulation of cytokines, cytokine inhibitors, and acute-phase proteins following anti-TNF-alpha therapy in rheumatoid arthritis. J Immunol 163(3):1521–1528
- 149. Nathan C (1987) Secretory products of macrophages. J Clin Invest 79:319–326
- Le JM, Weinstein D, Gubler U, Vilcek J (1987) Induction of membrane-associated interleukin 1 by tumor necrosis factor in human fibroblasts. J Immunol 138(7):2137–2142
- Nawroth PP, Bank I, Handley D, Cassimeris J, Chess L, Stern D (1986) Tumor necrosis factor/cachectin interacts with endothelial cell receptors to induce release of interleukin 1. J Exp Med 163(6): 1363–1375
- Philip R, Epstein LB (1986) Tumour necrosis factor as immunomodulator and mediator of monocyte cytotoxicity induced by itself, gamma-interferon and interleukin-1. Nature 323(6083):86– 89. doi:10.1038/323086a0
- Stashenko P, Dewhirst FE, Peros WJ, Kent RL, Ago JM (1987) Synergistic interactions between interleukin 1, tumor necrosis factor, and lymphotoxin in bone resorption. J Immunol 138(5):1464– 1468
- 154. Elias JA, Gustilo K, Freundlich B (1988) Human alveolar macrophage and blood monocyte inhibition of fibroblast proliferation. Evidence for synergy between interleukin-1 and tumor necrosis factor. Am Rev Respir Dis 138(6):1595–1603. doi:10.1164/ ajrccm/138.6.1595
- 155. Ogawa H, Nielsen S, Kawakami M (1989) Cachectin/tumor necrosis factor and interleukin-1 show different modes of combined effect on lipoprotein lipase activity and intracellular lipolysis in 3T3-L1 cells. Biochim Biophys Acta 1003(2):131–135

- 156. Bluthe RM, Pawlowski M, Suarez S, Parnet P, Pittman Q, Kelley KW, Dantzer R (1994) Synergy between tumor necrosis factor alpha and interleukin-1 in the induction of sickness behavior in mice. Psychoneuroendocrinology 19(2):197–207
- Rockett KA, Awburn MM, Rockett EJ, Clark IA (1994) Tumor necrosis factor and interleukin-1 synergy in the context of malaria pathology. AmJTrop Med Hyg 50(6):735–742
- 158. Rondeau JM, Ramage P, Zurini M, Gram H (2015) The molecular mode of action and species specificity of canakinumab, a human monoclonal antibody neutralizing IL-1beta. MAbs 7(6):1151–1160. doi:10.1080/19420862.2015.1081323
- 159. Goh AX, Bertin-Maghit S, Ping Yeo S, Ho AW, Derks H, Mortellaro A, Wang CI (2014) A novel human anti-interleukinlbeta neutralizing monoclonal antibody showing in vivo efficacy. MAbs 6(3):765–773. doi:10.4161/mabs.28614
- Seckinger P, Lowenthal JW, Williamson K, Dayer JM, MacDonald HR (1987) A urine inhibitor of interleukin 1 activity that blocks ligand binding. J Immunol 139(5):1546–1549
- Arend WP (1993) Interleukin-1 receptor antagonist. Adv Immunol 54:167–227
- Dinarello CA, van der Meer JW (2013) Treating inflammation by blocking interleukin-1 in humans. Semin Immunol 25(6):469– 484. doi:10.1016/j.smim.2013.10.008
- Loddick SA, Wong ML, Bongiorno PB, Gold PW, Licinio J, Rothwell NJ (1997) Endogenous interleukin-1 receptor antagonist is neuroprotective. Biochem Biophys Res Commun 234(1):211– 215. doi:10.1006/bbrc.1997.6436
- 164. Greenhalgh AD, Galea J, Denes A, Tyrrell PJ, Rothwell NJ (2010) Rapid brain penetration of interleukin-1 receptor antagonist in rat cerebral ischaemia: pharmacokinetics, distribution, protection. Br J Pharmacol 160(1):153–159. doi:10.1111/j.1476-5381.2010. 00684.x
- 165. Galea J, Ogungbenro K, Hulme S, Greenhalgh A, Aarons L, Scarth S, Hutchinson P, Grainger S, King A, Hopkins SJ, Rothwell N, Tyrrell P (2011) Intravenous anakinra can achieve experimentally effective concentrations in the central nervous system within a therapeutic time window: results of a dose-ranging study. J Cereb Blood Flow Metab 31(2):439–447. doi:10.1038/ jcbfm.2010.103
- 166. Singh N, Hopkins SJ, Hulme S, Galea JP, Hoadley M, Vail A, Hutchinson PJ, Grainger S, Rothwell NJ, King AT, Tyrrell PJ (2014) The effect of intravenous interleukin-1 receptor antagonist on inflammatory mediators in cerebrospinal fluid after subarachnoid haemorrhage: a phase II randomised controlled trial. J Neuroinflammation 11:1. doi:10.1186/1742-2094-11-1
- 167. Savman K, Blennow M, Hagberg H, Tarkowski E, Thoresen M, Whitelaw A (2002) Cytokine response in cerebrospinal fluid from preterm infants with posthaemorrhagic ventricular dilatation. Acta paediatrica (Oslo, Norway : 1992) 91(12):1357–1363
- 168. Rajatileka S, Odd D, Robinson MT, Spittle AC, Dwomoh L, Williams M, Harding D, Wagstaff M, Owen M, Crosby C, Ching J, Molnar E, Luyt K, Varadi A (2017) Variants of the EAAT2 glutamate transporter Gene promoter are associated with cerebral palsy in preterm infants. Mol Neurobiol. doi:10.1007/ s12035-017-0462-1